# THE LANCET Global Health

# Supplementary appendix

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#### **Methods**

**Table S1.** Real time PCR assays on TaqMan Array Card used for MAL-ED diarrhoea aetiology and growth analysis.

All the assays have been described previously and extensively validated.  $^{1-3}$  Nucleic acid was extracted with the QIAamp Fast DNA Stool mini kit (Qiagen, Hilden, Germany) with pre-treatment steps that included bead beating. AgPath One Step RT-PCR reagents were used for qPCR reactions, which were performed on ViiA 7, or QuantStudio 7, or QuantStudio 12K Flex systems. Quantification cycles (Cqs) are the PCR cycle values at which fluorescence from amplification exceeds the background, which acts as an inverse metric of quantity of nucleic acid. Valid results required proper functioning of controls (the negative results of a sample are valid only when its external control MS2 is positive,  $Cq \le 35$ ; the positive results are valid only when the corresponding extraction blank is negative for the relevant targets, Cq > 35), and excluded data flagged by the real time PCR software, i.e. BADROX in combination with NOISE or SPIKE.

	Pathogen	Gene	Included in diarrhoea- growth analysis	Included in infection- growth analysis	Assay in the original microbiologic work-up (conventional methods)*
Viruses	Adenovirus 40/41	Fiber gene	✓	✓	EIA <sup>†</sup>
	Astrovirus	Capsid	✓	✓	EIA
	Norovirus GI/GII	GI ORF1-2 and GII ORF1-2	✓	✓	PCR <sup>§</sup>
	Rotavirus	NSP3	✓		EIA
	Sapovirus	RdRp	✓	✓	Not tested
Bacteria	EAEC**	aaiĈ, aatA, aggR	✓	✓	Culture/PCR
	Atypical EPEC**	eae	✓	✓	Culture/PCR
	Typical EPEC**	bfpA	✓	✓	Culture/PCR
	ETEC**	LT, $STh$ and $STp$	✓	✓	Culture/PCR
	STEC**	stx1, stx2	✓		Culture/PCR
	Aeromonas	Aerolysin	✓		Culture
	Campylobacter spp.	cadF (C. jejuni/coli) and cpn60 (Campylobacter spp)	✓ (C. jejuni/coli)	✓ (Campylobacter spp)	$ extsf{EIA}^{\dagger\dagger}$
	Helicobacter pylori	ureC	✓		Not tested
	Plesiomonas shigelloides	gyrB	✓		Culture
	Salmonella	ttr	✓		Culture
	Shigella/EIEC	ipaH	✓	✓	Culture
	Vibrio cholerae	ĥlyA	✓		Culture
Fungi	Enterocytozoon bieneusi	ITS	✓		Not tested
	Encephalitozoon intestinalis	SSU rRNA	✓		Not tested
Protozoa	Cryptosporidium	18S rRNA	✓	✓	EIA
	Cyclospora cayetanensis	18S rRNA	✓		Microscopy
	Cystoisospora belli	18S rRNA	✓		Microscopy
	Entamoeba histolytica	18S rRNA	✓		EIA
	Giardia	18S rRNA	✓	✓	EIA
Helminth	Ancylostoma duodenale	ITS2	✓		Microscopy
	Ascaris lumbricoides	ITS1			Microscopy
	Necator americanus	ITS2			Microscopy
	Strongyloides stercoralis	Dispersed repetitive sequence	✓		Microscopy
	Trichuris trichiura	18S rRNA	✓		Microscopy
Controls	MS2	MS2g1			N/A
	PhHV	gB °			N/A

<sup>\*</sup> Conventional methods were conducted in non-diarrhoeal stools monthly in the first year of life, but only quarterly in the second year of life (qPCR was conducted in non-diarrhoeal stools monthly for both years).

<sup>†</sup> Pan-adenovirus EIA only.

<sup>§</sup> Norovirus PCR was only performed on diarrhoeal and non-diarrhoeal stools from a randomly selected 10% of participants.

<sup>\*\*</sup>E. coli pathotypes were defined as follows: EAEC (aaiC, or aatA, or both), atypical EPEC (eae without bfpA, stx1, and stx2), typical EPEC (bfpA), ETEC (STh, STp, or LT), STEC (eae without bfpA and with stx1, stx2, or both). EAEC was alternatively defined by aggR in sensitivity analysis.

<sup>††</sup>A single EIA was used, which has been shown to detect some Campylobacter species other than C. jejuni and C. coli. 4

#### **Longitudinal model methods and validation**

All analyses were conducted in SAS version 9.4 (Cary, North Carolina, USA). To implement the parametric g-formula, we first used the observed data to estimate  $\beta$ -coefficients in a series of longitudinal repeated measures models for each time-varying covariate in the 6-month intervals. Results from models with 3-month and 6-month intervals were largely consistent; we present the 6-month interval results because 3-month interval-based estimates were less precise and more sensitive to model specification. All models included an indicator for the interval, LAZ at the beginning of the interval, site, sex, and SES as covariates. The time-varying covariates modelled were:

- 1. Percent days with exclusive breastfeeding: modelled with Poisson regression with an offset for the number of days under surveillance in the period, and also adjusting for maternal age.
- 2. Incidence of diarrhoea (number of episodes): modelled with Poisson regression and also adjusting for exclusive breastfeeding, and detection of *Campylobacter*, *Shigella*, and tEPEC in the previous interval.
- 3. Macrolide use (yes/no): modelled with logistic regression and also adjusting for exposure to EAEC, *Campylobacter*, *Giardia*, *Shigella*, and tEPEC in the previous interval, number of diarrhoea episodes, exclusive breastfeeding, and improved sanitation. Macrolide use was included since it was a strong predictor of *Campylobacter*, *Shigella*, and tEPEC infection.
- 4. The number of detections of pathogens that were not the exposure of interest (*Campylobacter* spp., *Giardia*, EAEC, tEPEC, and *Shigella*): modelled with Poisson regression with an offset for the number of stool samples tested for that pathogen, and also adjusting for exposure to that pathogen in the previous interval, exposure to the other four pathogens in the previous interval, exclusive breastfeeding, number of diarrhoea episodes, macrolide use, maternal height, improved sanitation, and improved drinking water.
- 5. The average quantity of these pathogens: modelled with the same covariates using log-linear regression only among those with positive detections.
- 6. The number of detections or average quantity of the pathogen of interest: similarly modelled with Poisson and log-linear regression, respectively, but further adjusted for exposure to the other pathogens (from step 4) in the current interval.
- 7. The outcome, LAZ, at the end of the interval: modelled using a separate linear regression model for each 6-month time period, and additionally adjusted for the current and lagged pathogen exposure variables (no lags during the first 6-month period, and progressively more lags for later periods), exclusive breastfeeding (first period only), and maternal height (Figure S1).

These time-varying covariates were considered for inclusion based on a causal directed acyclic graph. Covariates were dropped from models for specific pathogen detections if they did not have a strong (|LAZ| difference |>0.02) or significant association (p<0.05) with that pathogen and if model fit improved by Akaike information criterion (AIC).

We then used Monte Carlo simulations with the estimated coefficients from the models above to predict the time-varying covariates, pathogen exposures, and LAZ outcomes for each interval to 2 years of age in a random sample of 50,000 replicates from the study population at baseline. The simulation was completed for the natural course scenario, in which all pathogen exposures were predicted based on the observed data, to validate model fit. The simulated data under the natural course scenario fit the observed data well, such that the interval-specific distributions of the time-varying covariates and LAZ outcomes were similar between the observed and simulated data (Figure S2).

The simulations were then completed for high and low pathogen exposure scenarios, which produced the predicted covariates and outcomes had all children been exposed to high pathogen exposure, defined by the 90<sup>th</sup> percentile in each interval, versus had all children been exposed to low pathogen exposure, defined by the 10<sup>th</sup> percentile in each interval. To estimate the cumulative effect of pathogen exposure on LAZ at 2 years of age, we averaged the predicted LAZ outcomes at 2 years across individuals in the high and low pathogen exposure scenarios and took their difference to estimate the population-standardized LAZ difference. A similar process was used to estimate the cumulative effect of pathogen exposure on LAZ at 2 years of age per one log increase in pathogen quantity. Confidence intervals were constructed by bootstrap of the above steps with 1000 replicates.

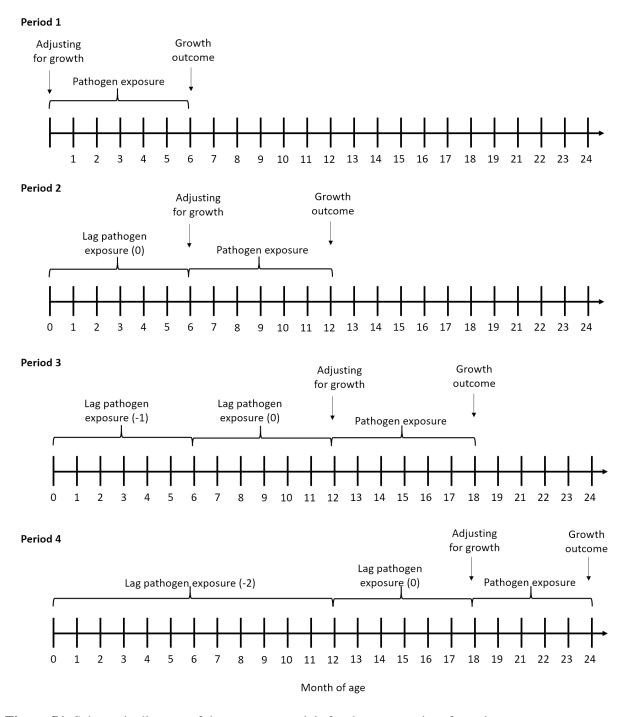
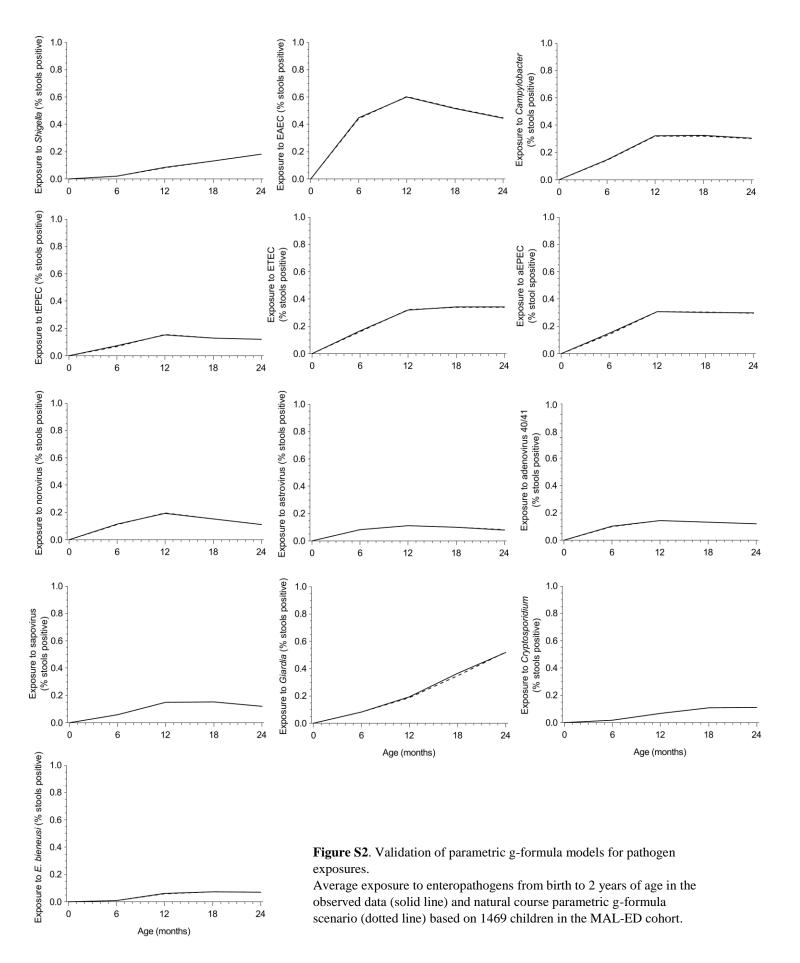
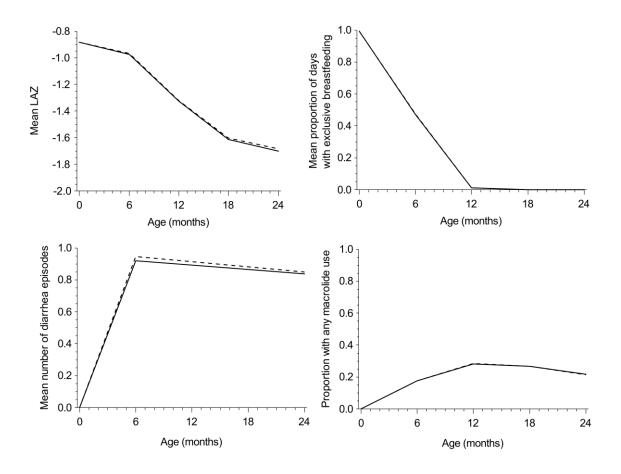


Figure S1. Schematic diagram of the outcome models for the parametric g-formula.





**Figure S3**. Validation of parametric g-formula models for time-varying covariates and LAZ outcomes. Average LAZ, proportion of days with exclusive breastfeeding, number of diarrhoea episodes, and macrolide use from birth to 2 years of age in the observed data (solid line) and natural course parametric g-formula scenario (dotted line) based on 1469 children in the MAL-ED cohort.

## **Enteropathogen prevalence and quantity**

Table S2. Enteropathogen prevalence in non-diarrhoeal stools during the first 2 years of life by site among 1469 children in the MAL-ED cohort.

			Percent non-diarrhoeal stools positive N (%)						
			Dhaka,	Vellore,	Bhaktapur,	Fortaleza,		Venda,	Haydom,
	Site	All	Bangladesh	India	Nepal	Brazil	Loreto, Peru	South Africa	Tanzania
Bacteria	Shigella	3237 (10.8)	564 (13·1)	592 (12.4)	290 (5.8)	139 (4.9)	574 (13.6)	321 (7.0)	757 (17.8)
	EAEC	15314 (51.0)	1877 (43.6)	3052 (63.9)	2503 (49.6)	857 (30·1)	2621 (62-2)	1654 (35.9)	2750 (65.0)
	Campylobacter	8398 (28.0)	1702 (39.4)	1177 (24.7)	1325 (26.3)	428 (15.0)	1044 (24.8)	525 (11.4)	2197 (51.8)
	tEPEC	3653 (12.2)	862 (20.0)	770 (16.2)	344 (6.8)	97 (3.4)	590 (14.0)	201 (4.4)	789 (18.6)
	ETEC	9056 (30.0)	2026 (46.8)	1471 (30.7)	1057 (20.9)	222 (7.7)	1349 (32.0)	578 (12.6)	2353 (54.8)
	aEPEC	8054 (26.8)	1083 (25.1)	1339 (28·1)	1535 (30.4)	742 (25.9)	1203 (28.6)	924 (20.2)	1228 (28.8)
Viruses	Norovirus	4338 (14.5)	836 (19.4)	730 (15.4)	628 (12.4)	161 (5.7)	806 (19·1)	477 (10.5)	700 (16.5)
	Astrovirus	2862 (9.6)	792 (18.3)	492 (10.4)	253 (5.0)	45 (1.6)	695 (16.6)	313 (6.8)	272 (6.4)
	Adenovirus 40/41	3791 (12.7)	779 (18.0)	787 (16.5)	296 (5.9)	126 (4.4)	969 (23.3)	491 (10.7)	343 (8.1)
	Sapovirus	3680 (12.3)	746 (17.3)	662 (13.9)	536 (10.6)	122(4.3)	660 (15.7)	501 (10.9)	453 (10.7)
Protozoa	Giardia	8898 (29.7)	923 (21-4)	1804 (37.6)	1036 (20.5)	442 (15.5)	1760 (42.3)	1099 (23.6)	1834 (43.8)
	Cryptosporidium	2363 (7.9)	364 (8.4)	257 (5.4)	318 (6.3)	38 (1.3)	528 (12.5)	250 (5.5)	608 (14.3)
	E. bieneusi*	1674 (5.6)	354 (8.2)	135 (2.8)	161 (3.2)	19 (0.7)	256 (6.1)	190 (4.2)	559 (13.2)

<sup>\*</sup>E. bieneusi is an intracellular parasitic fungus

**Table S3**. Enteropathogen quantity in positive non-diarrhoeal stools during the first 2 years of life by site among 1469 children in the MAL-ED cohort.

Mean quantity in log (copy number) per gram of stool (standard deviation) Dhaka, Haydom, Vellore, Bhaktapur, Fortaleza, Loreto, Venda, Bangladesh **Brazil** Tanzania Site All India Nepal Peru South Africa Bacteria Shigella 5.62(1.29)5.61(1.19)5.47(1.20)5.64(1.26)5.14(1.24)5.81 (1.43) 5.05(0.94)5.91 (1.37) **EAEC** 6.12 (1.52) 5.97 (1.56) 6.06(1.46)6.59(1.46)5.66(1.37)6.48(1.65)5.35(1.09)6.14(1.52)Campylobacter 4.91(1.14)5.74 (1.33) 5.95 (1.36) 5.37(1.21)6.06(1.41)4.88(0.85)5.85 (1.37) 5.91 (1.21) tEPEC 5.83 (1.43) 5.79 (1.49) 6.28(1.34)5.38(1.18)5.88 (1.55) 5.37(1.08)5.92 (1.41) 5.71(1.37)**ETEC** 5.88 (1.60) 5.68 (1.56) 5.65(1.49)6.47(1.64)5.47(1.38)5.89(1.73)5.27(1.17)6.12(1.59)aEPEC 5.45 (1.24) 5.46 (1.26) 5.35 (1.19) 5.79(1.23)5.30(1.13)5.70(1.37)4.88(0.91)5.40(1.25)Viruses Norovirus 5.25 (1.10) 5.29 (1.10) 5.18 (1.03) 5.27(1.14)5.01(1.00)5.41 (1.14) 5.05(1.03)5.29(1.10)Astrovirus 6.17(1.99)5.64(1.93)5.22 (1.68) 5.15 (1.55) 5.37 (1.71) 5.25 (1.55) 5.17(1.55)5.89(2.00)Adenovirus 40/41 5.35 (1.64) 5.28 (1.44) 6.10(2.36)5.55(1.61)5.22(1.45)5.00(1.35)5.17(1.47)6.08(2.08)Sapovirus 5.55 (1.50) 5.63 (1.52) 5.37(1.44)5.69(1.61)5.53(1.40)5.50(1.52)5.38(1.39)5.83(1.46)Protozoa Giardia 6.36(1.77) 5.72(1.66)5.98 (1.63) 7.54(1.74)5.92(1.32)6.63(1.78)5.40(1.45)6.80(1.67)Cryptosporidium 5.41 (1.34) 5.46 (1.47) 5.21(1.13)5.52(1.37)4.95(1.08)5.64(1.47)5.06 (1.15) 5.39(1.25)E. bieneusi\* 5.63 (1.39) 5.87 (1.35) 5.62 (1.53) 5.54(1.34)5.68 (1.46) 5.67(1.45)5.45(1.18)5.37(1.30)

<sup>\*</sup>E. bieneusi is an intracellular parasitic fungus

**Table S4**. Enteropathogen prevalence per child in 6 month intervals during the first 2 years of life among 1469 children in the MAL-ED cohort.

		Median percent nor	Median percent non-diarrhoeal stools positive per child (10 <sup>th</sup> , 90 <sup>th</sup> percentiles)					
	Months	1-24	1-6	7-12	13-18	19-24		
Bacteria	Shigella	0.09 (0-0.24)	0 (0-0)	0 (0-0.33)	0 (0-0.40)	0.17 (0-0.50)		
	EAEC	0.50(0.25-0.74)	0.4(0-1)	0.6(0.20-1)	0.5 (0.17 - 0.83)	0.5 (0-0.80)		
	Campylobacter	0.24 (0.05 - 0.57)	0(0-0.50)	0.33(0-0.75)	0.25(0-0.80)	0.2(0-0.80)		
	tEPEC	0.10(0-0.25)	0(0-0.25)	0.17(0-0.40)	0(0-0.33)	0(0-0.33)		
	ETEC	0.27 (0.06 - 0.57)	0.17(0-0.5)	0.25(0-0.75)	0.33(0-0.75)	0.33(0-0.75)		
	aEPEC	0.26(0.13-0.41)	0(0-0.40)	0.33(0-0.60)	0.33(0-0.60)	0.33 (0-0.60)		
Viruses	Norovirus	1.14 (0.04 - 0.26)	0(0-0.33)	0.17(0-0.50)	0.17(0-0.40)	0(0-0.33)		
	Astrovirus	0.08(0-0.22)	0(0-0.33)	0(0-0.33)	0(0-0.33)	0(0-0.25)		
	Adenovirus 40/41	0.10(0-0.27)	0(0-0.33)	0(0-0.40)	0(0-0.33)	0(0-0.33)		
	Sapovirus	0.11 (0-0.23)	0 (0-0.20)	0.17 (0-0.40)	0.17 (0-0.40)	0 (0-0.33)		
Protozoa	Giardia	0.25(0-0.63)	0 (0-0.33)	0(0-0.67)	0.2(0-1)	0.6 (0-1)		
	Cryptosporidium	0.05 (0-0.18)	0 (0–0)	0(0-0.25)	0 (0-0.33)	0 (0-0.33)		
	E. bieneusi*	0 (0-0.16)	0 (0-0)	0 (0-0.33)	0 (0-0.33)	0 (0-0.33)		

<sup>\*</sup>E. bieneusi is an intracellular parasitic fungus

**Table S5**. Enteropathogen quantity per child during the first 2 years of life among 1469 children in the MAL-ED cohort.

		Median quantity in	log (copy number	r) per gram of stool (	10 <sup>th</sup> , 90 <sup>th</sup> percentiles)	
	Months	1-24	1-6	7-12	13-18	19-24
Bacteria	Shigella	0.49 (0-1.39)	0 (0-0)	0 (0-1.85)	0 (0-2·27)	0.74 (0-3.02)
	EAEC	3.12(1.37-4.69)	2.71(0-6.28)	3.74(0.98-6.39)	3.01 (0.82 - 5.44)	2.46 (0-4.71)
	Campylobacter	1.32(0.27-3.30)	0(0-2.78)	1.60(0-4.35)	1.39(0-4.52)	1.12(0-4.21)
	tEPEC	0.62(0-1.46)	0(0-1.75)	0.72(0-2.50)	$0(0-2\cdot18)$	0(0-1.99)
	ETEC	1.63(0.31-3.38)	0 (0–0)	1.56(0-3.51)	1.52(0-3.30)	1.39 (0-3.34)
	aEPEC	1.39(0.63-2.30)	0(0-1.32)	0.70(0-2.23)	0.75(0-2.17)	0(0-1.89)
Viruses	Norovirus	0.71(0.18-1.39)	0(0-2.05)	0.92(0-2.52)	0.75(0-1.96)	0(0-1.56)
	Astrovirus	0.43(0-1.17)	0(0-1.50)	0(0-1.86)	0(0-1.74)	0(0-1.49)
	Adenovirus 40/41	0.56(0-1.44)	0(0-1.72)	$0(0-2\cdot22)$	$0(0-2\cdot03)$	0(0-1.92)
	Sapovirus	0.62(0-1.27)	0 (0-0)	0(0-1.48)	0(0-1.94)	0(0-1.85)
Protozoa	Giardia	1.45 (0-4.28)	0(0-1.40)	0(0-3.95)	1.22 (0-6.77)	3.38 (0-7.40)
	Cryptosporidium	0.32(0-0.99)	0(0-1.84)	0.70(0-2.57)	0.72(0-2.41)	0.72(0-2.16)
	E. bieneusi*	0(0-0.89)	0 (0–0)	0 (0–1.65)	0 (0–1.59)	0 (0–1.66)

<sup>\*</sup>E. bieneusi is an intracellular parasitic fungus

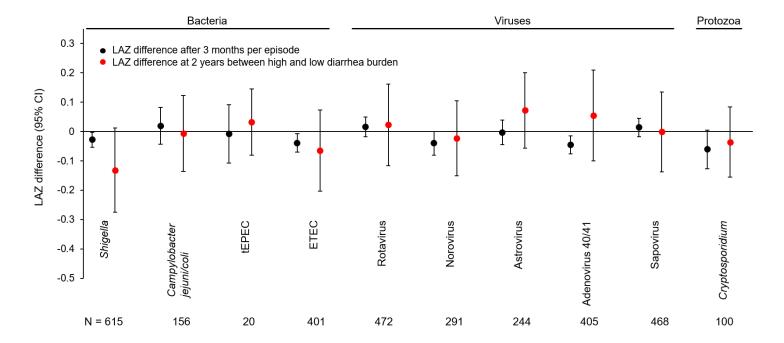


Figure S4. Associations of pathogen-attributable diarrhoea with length-for-age z-score.

Per-episode association of diarrhoea with length-for-age z-score (LAZ) 3 months after the episode (LAZ difference and 95% CI; black dots) and LAZ difference and 95% CI at 2 years of age between high (90<sup>th</sup> percentile) and low (10<sup>th</sup> percentile) pathogen-attributable diarrhoea burden (red dots). Analysis includes 37951 observed child-months among 1469 children in the MAL-ED cohort with molecular testing of stool samples. Per-episode estimates were adjusted for age, site, sex, SES, maternal height, LAZ at the beginning of the interval, exclusive breastfeeding, and non-attributable diarrhoea episodes in the same period. Two-year estimates were adjusted for site, sex, SES, maternal height, enrolment LAZ, exclusive breastfeeding in the first 6 months of life, number of antibiotic courses, and number of non-attributable diarrhoea episodes.

## **Site-specific effects**

**Table S6.** Site-specific associations of pathogen-attributable diarrhoea episodes on LAZ 3 months following the episode adjusted for age, sex, SES, maternal height, and antibiotic treatment of the episode among 1469 children in the MAL-ED cohort.

	Diarrhoea aetiology					
Site	Any infectious aetiology	Bacterial*	Viral <sup>†</sup>	Parasitic <sup>‡</sup>		
Dhaka, Bangladesh	-0.06 (-0.08 to -0.03)	-0.05 (-0.08 to -0.02)	-0.03 (-0.06-0.00)	0.01 (-0.10-0.11)		
Vellore, India	-0.04 (-0.07  to  -0.01)	-0.02 (-0.06-0.02)	-0.03 (-0.07-0.01)	-0.04 (-0.17-0.10)		
Bhaktapur, Nepal	0.02 (-0.01-0.05)	0.00 (-0.05-0.05)	0.05 (0.01-0.08)	-0·14 (-0·26 to -0·02)		
Fortaleza, Brazil	-0.10 (-0.23-0.02)	-0.03 (-0.19-0.12)	-0.10 (-0.27-0.07)	-0.27 (-0.60–0.05)		
Loreto, Peru	-0.02 (-0.05-0.01)	-0.05 (-0.09  to  -0.01)	0.01 (-0.02-0.05)	-0.10 (-0.19-0.00)		
Venda, South Africa	0.05 (-0.06-0.17)	0.03 (-0.16-0.22)	0.05 (-0.10-0.19)	$N/A^{\S}$		
Haydom, Tanzania	-0.13 (-0.22  to  -0.04)	-0.04 (-0.18-0.09)	-0.15 (-0.28  to  -0.01)	-0.15 (-0.41–0.11)		

<sup>\*</sup> Includes EAEC, aEPEC, tEPEC, ETEC, STEC, Aeromonas, C. jejuni/coli, H. pylori, Plesiomonas, Salmonella, Shigella, and V. cholera

<sup>†</sup> Includes adenovirus 40/41, astrovirus, norovirus GI/GII, rotavirus, sapovirus

<sup>‡</sup> Includes E. bieneusi, E. intestinalis, Cryptosporidium, Cyclospora, Isospora, E. histolytica, Giardia, Ancylostoma, Strongyloides, and Trichuris

<sup>§</sup> No parasite-attributable diarrhoea episodes were identified in Venda, South Africa

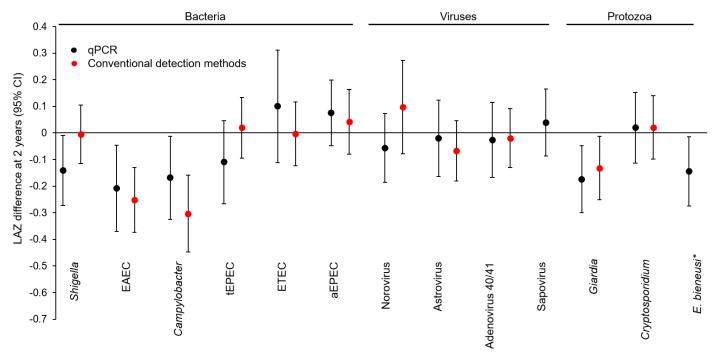
**Table S7**. Site-specific differences in length-for-age z-score at 2 years of age associated with the difference between high (site-specific 90<sup>th</sup> percentile) and low (site-specific 10<sup>th</sup> percentile) exposure to each of the 13 most prevalent enteropathogens in non-diarrhoeal stools among 1469 children in the MAL-ED cohort.

		All sites	Dhaka, Bangladesh	Vellore, India	Bhaktapur, Nepal
Bacteria	Shigella	-0.14 (-0.27  to  -0.01)	-0.23 (-0.53–0.08)	0.00 (-0.28-0.28)	0.21 (-0.04-0.46)
	EAEC	-0·21 (-0·37 to -0·05)	-0.26 (-0.52–0.01)	0.11 (-0.16-0.39)	0.07 (-0.19-0.32)
	Campylobacter	-0·17 (-0·32 to -0·01)	-0.19 (-0.50-0.13)	-0.05 (-0.33-0.24)	-0·34 (-0·61 to -0·07)
	tEPEC	-0.11 (-0.27–0.05)	0.15 (-0.14-0.44)	-0.10 (-0.38-0.18)	-0·38 (-0·64 to -0·11)
	ETEC	0.10 (-0.11-0.31)	0.01 (-0.29-0.31)	-0.07 (-0.35–0.21)	0.04 (-0.23-0.30)
	aEPEC	0.08 (-0.05-0.20)	-0.03 (-0.31–0.25)	-0.18 (-0.47-0.11)	0.14 (-0.12-0.40)
Viruses	Norovirus	-0.06 (-0.19-0.07)	-0.25 (-0.53–0.02)	-0.26 (-0.53-0.01)	0.17 (-0.08-0.42)
	Astrovirus	-0.02 (-0.16-0.12)	-0.04 (-0.31–0.23)	-0.08 (-0.35-0.19)	0.02 (-0.23-0.28)
	Adenovirus 40/41	-0.03 (-0.17-0.11)	-0.08 (-0.35–0.19)	0.12 (-0.14-0.38)	0.14 (-0.10-0.38)
	Sapovirus	0.04 (-0.09-0.16)	-0.15 (-0.43–0.13)	0.18 (-0.09-0.44)	0.16 (-0.09-0.41)
Protozoa	Giardia	-0.17 (-0.30  to  -0.05)	0.09 (-0.18-0.36)	-0.25 (-0.52–0.03)	-0.18 (-0.43-0.06)
	Cryptosporidium	0.02 (-0.11-0.15)	0.23 (-0.04-0.51)	-0.04 (-0.30-0.22)	-0.18 (-0.45–0.09)
	E∙ bieneusi*	-0·14 (-0·27 to -0·01)	0.09 (-0.19-0.38)	-0.05 (-0.32–0.21)	-0.15 (-0.39–0.10)

		Fortaleza, Brazil	Loreto, Peru	Venda, South Africa	Haydom, Tanzania
Bacteria	Shigella	-0.03 (-0.48-0.42)	-0.23 (-0.51–0.04)	-0.23 (-0.55–0.08)	-0·36 (-0·67 to -0·04)
	EAEC	-0.28 (-0.72–0.16)	-0.04 (-0.31–0.23)	-0.34 (-0.67–0.00)	-0.40 (-0.72  to  -0.07)
	Campylobacter	-0.01 (-0.4-0.38)	0.08 (-0.19-0.34)	-0.30 (-0.61–0.01)	-0.06 (-0.37-0.24)
	tEPEC	0.00 (-0.39-0.39)	0.09 (-0.19-0.37)	-0.11 (-0.42-0.20)	-0.03 (-0.36–0.31)
	ETEC	0.14 (-0.28–0.56)	-0.17 (-0.45–0.11)	0.11 (-0.22-0.44)	0.38 (0.06–0.71)
	aEPEC	-0.17 (-0.59–0.26)	0.31 (0.03-0.60)	0.13 (-0.18-0.45)	0.31 (-0.01–0.63)
Viruses	Norovirus	0.06 (-0.35, 0.47)	-0.18 (-0.46–0.10)	0.03 (-0.29-0.35)	0.18 (-0.12-0.47)
	Astrovirus	-0.55 (-0.94 to -0.16)	-0.13 (-0.39-0.12)	0.16 (-0.14-0.47)	0.20 (-0.09-0.48)
	Adenovirus 40/41	-0.01 (-0.43–0.41)	-0.14 (-0.41–0.13)	-0.19 (-0.49-0.11)	-0.04 (-0.32–0.25)
	Sapovirus	0.33 (-0.09-0.75)	0.16 (-0.10-0.43)	0.05 (-0.26-0.36)	-0.19 (-0.48-0.10)
Protozoa	Giardia	0.16 (-0.28–0.61)	-0·33 (-0·60 to -0·05)	-0·35 (-0·66 to -0·04)	-0.10 (-0.40-0.21)
	Cryptosporidium	-0.03 (-0.43–0.37)	0.03 (-0.23-0.29)	0.38 (0.06–0.69)	-0.07 (-0.37–0.23)
	E∙ bieneusi*	-0.06 (-0.46–0.34)	-0.19 (-0.46–0.07)	-0.24 (-0.55–0.07)	-0.24 (-0.54–0.05)

<sup>\*</sup>E. bieneusi is an intracellular parasitic fungus

### Height attainment model sensitivity analyses

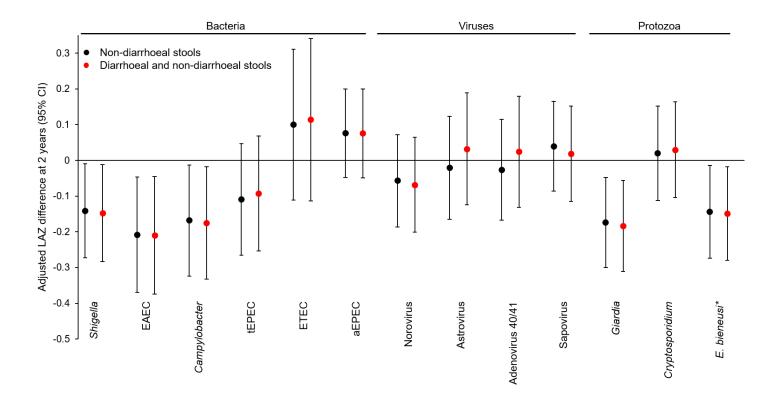


**Figure S5**. Height attainment model: qPCR vs. conventional methods.

Difference and 95% confidence interval in length-for-age z-score (LAZ) at 2 years of age associated with the difference between high (90<sup>th</sup> percentile) and low (10<sup>th</sup> percentile) pathogen prevalence for each of the 13 most prevalent enteropathogens using quantitative PCR detection methods (black) and conventional detection methods (Table S1; red) in non-diarrhoeal stools among 1469 children in the MAL-ED cohort. Estimates are adjusted for site, enrolment LAZ, sex, SES, exclusive breastfeeding in the first 6 months of life, and maternal height.

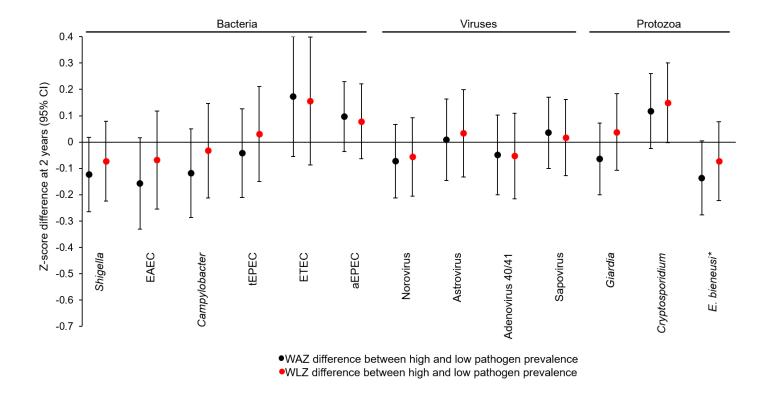
Note: Sapovirus and *E. bieneusi* were not assayed using conventional methods and norovirus was only tested using conventional methods in random 10% of children. Conventional methods were conducted in non-diarrhoeal stools monthly in the first year of life and only quarterly in the second year, while qPCR was conducted in non-diarrhoeal stools monthly in both years.

<sup>\*</sup>E. bieneusi is an intracellular parasitic fungus



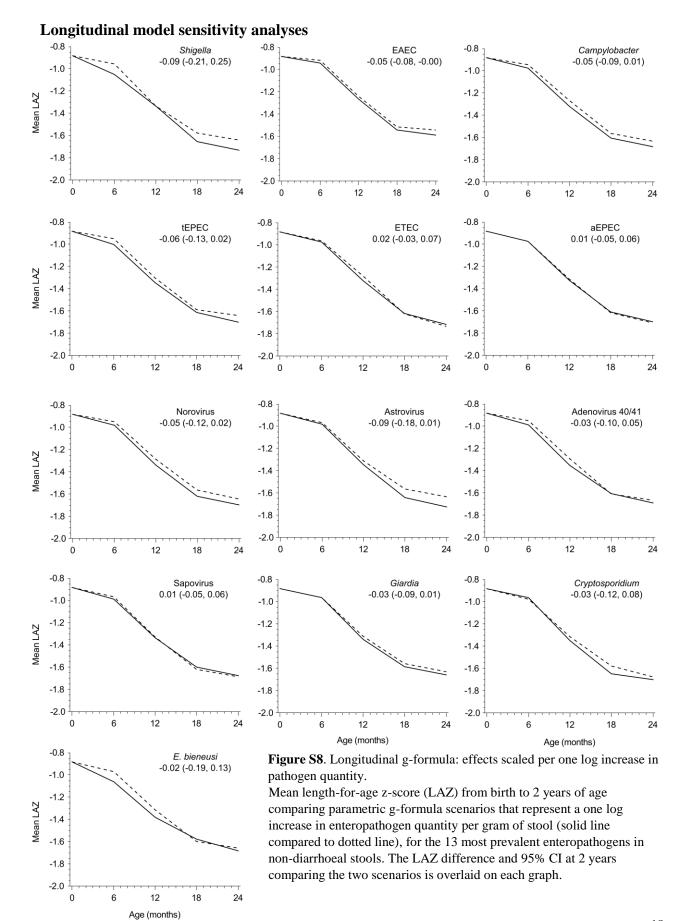
**Figure S6.** Height attainment model: non-diarrhoeal stools and diarrhoeal stools. Difference and 95% confidence interval in LAZ at 2 years of age associated with the difference between high (90<sup>th</sup> percentile) and low (10<sup>th</sup> percentile) pathogen prevalence for each of the 13 most prevalent enteropathogens in non-diarrhoeal stools (black) and all stools (diarrhoeal and non-diarrhoeal; red) among 1469 children in the MAL-ED cohort. Estimates are adjusted for site, enrolment WAZ and LAZ, sex, SES, exclusive breastfeeding in the first 6 months of life, and maternal height.

<sup>\*</sup>E. bieneusi is an intracellular parasitic fungus



**Figure S7.** Height attainment model: weight-for-age (WAZ) and weight-for-length (WLZ) z-scores. Difference and 95% confidence interval in WAZ and WLZ at 2 years of age associated with the difference between high (90<sup>th</sup> percentile) and low (10<sup>th</sup> percentile) pathogen prevalence for each of the 13 most prevalent enteropathogens in non-diarrhoeal stools among 1469 children in the MAL-ED cohort. Estimates are adjusted for site, enrolment WAZ and LAZ, sex, SES, exclusive breastfeeding in the first 6 months of life, and maternal height.

<sup>\*</sup>E. bieneusi is an intracellular parasitic fungus



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